

4-11-2018

Feeding reduced-fat dried distillers grains with solubles to lactating Holstein dairy cows does not alter milk composition or cause late blowing in cheese

E.D. Testroet
Washington State University

Donald C. Beitz
Iowa State University, dcbeitz@iastate.edu


M.R. O'Neil
Iowa State University

A. L. Mueller
Purina Animal Nutrition

H. A. Ramirez-Ramirez
Iowa State University

See next page for additional authors

Follow this and additional works at: https://lib.dr.iastate.edu/fshn_hs_pubs

 Part of the [Dairy Science Commons](#), [Food Chemistry Commons](#), and the [Molecular, Genetic, and Biochemical Nutrition Commons](#)

The complete bibliographic information for this item can be found at https://lib.dr.iastate.edu/fshn_hs_pubs/42. For information on how to cite this item, please visit <http://lib.dr.iastate.edu/howtocite.html>.

This Article is brought to you for free and open access by the Food Science and Human Nutrition at Iowa State University Digital Repository. It has been accepted for inclusion in Food Science and Human Nutrition Publications by an authorized administrator of Iowa State University Digital Repository. For more information, please contact digirep@iastate.edu.

Feeding reduced-fat dried distillers grains with solubles to lactating Holstein dairy cows does not alter milk composition or cause late blowing in cheese

Abstract

Feeding dried distillers grains with solubles (DDGS) to lactating dairy cows has been implicated as a cause of late blowing defects in the production of Swiss-style cheeses. Our objectives were (1) to test the effect of feeding reduced-fat DDGS (RF-DDGS; ~6% fat) to lactating dairy cows on the composition of milk and on the suitability of the milk for production of baby Swiss cheese and (2) to evaluate the effect of diet on cow lactation performance. Lactating Holstein dairy cows were fed both dietary treatments in a 2 × 2 crossover design. Cows were housed in a 48-cow freestall pen equipped with individual feeding gates to record feed intake. The control diet was a corn, corn silage, and alfalfa hay diet supplemented with mechanically expelled soybean meal. The experimental diet was the same base ration, but 20% (dry matter basis) RF-DDGS were included in place of the expelled soybean meal. The RF-DDGS diet was additionally supplemented with rumen-protected lysine; diets were formulated to be isoenergetic and isonitrogenous. Cows were allowed ad libitum access to feed and water, fed twice daily, and milked 3 times daily. For cheese production, milk was collected and pooled 6 times for each dietary treatment. There was no treatment effect on milk yield (35.66 and 35.39 kg/d), milk fat production (1.27 and 1.25 kg/d), milk fat percentage (3.65 and 3.61%), milk protein production (1.05 and 1.08 kg/d), lactose percentage (4.62 and 4.64%), milk total solids (12.19 and 12.28%), and somatic cell count (232.57 and 287.22 × 10³ cells/mL) for control and RF-DDGS, respectively. However, dry matter intake was increased by treatment, which implied a reduction in feed efficiency. Milk protein percentage also increased (3.01 and 3.11%), whereas milk urea nitrogen decreased (14.18 and 12.99 mg/dL), indicating that protein utilization may be more efficient when cows are fed RF-DDGS. No differences in cheese were observed by a trained panel except cheese appearance; control cheese eyes were significantly, but not practically, larger than the RF-DDGS cheese. These results indicate that RF-DDGS can be effectively used in the rations of lactating Holstein cows with no deleterious effects on milk production and composition and metrics of the physiology of the cow (i.e., blood glucose and nonesterified fatty acids); however, feeding RF-DDGS increased dry matter intake, which decreased feed efficiency. Finally, feeding RF-DDGS did not negatively influence quality and suitability of milk for production of baby Swiss cheese.

Keywords

Efficiency, dried distillers grains with solubles, milk fat, protein utilization, sensory

Disciplines

Dairy Science | Food Chemistry | Food Science | Molecular, Genetic, and Biochemical Nutrition

Comments

This accepted article is published as Testroet, E. D., D. C. Beitz, and S. Clark*. 2018. Feeding reduced-fat dried distillers grains with solubles to lactating Holstein dairy cows does not negatively influence cow performance or cheese quality. *J Dairy Sci.* 2018 Jul;101(7):5838-5850. doi: [10.3168/jds.2017-13699](https://doi.org/10.3168/jds.2017-13699). Posted with permission.

Authors

E.D. Testroet, Donald C. Beitz, M.R. O'Neil, A. L. Mueller, H. A. Ramirez-Ramirez, and Stephanie Clark

1 **Feeding reduced-fat dried distillers grains with solubles to lactating Holstein dairy cows**
2 **does not negatively influence cow performance or cause late blowing in cheese**

3 **E. D. Testroet**

4 **Interpretive summary**

5 Holstein dairy cows were fed a control diet not containing reduced-fat distillers grains with
6 solubles (RF-DDGS), or a diet that contained 20% of the dry matter (DM) as RF-DDGS. Milk
7 composition was not affected when cows were fed RF-DDGS, except for an increase in milk
8 protein percentage. Cows fed either diet performed equally well, but cows fed RF-DDGS had
9 improved nitrogen efficiency. These results show that cows can be fed 20% RF-DDGS without
10 a loss in performance. Furthermore, milk from cows fed RF-DDGS was used to produce baby
11 Swiss cheese that did not differ in quality from cheese made from milk of cows fed the control
12 diet.

13

14 **RUNNING HEAD: RF-DDGS, COW PERFORMANCE, AND CHEESE QUALITY**

15

16 **Feeding reduced-fat dried distillers grains with solubles to lactating Holstein dairy cows**
17 **does not negatively influence cow performance or cause late blowing in cheese**

18

19 **E. D. Testroet,* D. C. Beitz,† M. R. O’Neil,† A. L. Mueller,‡ H. A. Ramirez-Ramirez,† and**
20 **S. Clark*§**

21 * Department of Food Science and Human Nutrition, Iowa State University, Ames, IA 50011

22 † Department of Animal Science, Iowa State University, Ames, IA 50011

23 ‡ Land O’Lakes Purina, LLC, Pierz, MN, 563643

24 §Corresponding Author: S. Clark, milkmade@iastate.edu

25

26 **ABSTRACT**

27 Feeding dried distillers grains with solubles (DDGS) to lactating dairy cows has been implicated
28 as a cause of late blowing defects in production of Swiss-style cheeses. Our objectives were to
29 test the impact of feeding reduced-fat dried distillers grains with solubles (RF-DDGS; ~6% fat)
30 to lactating dairy cows on the composition of milk and on the suitability of the milk for
31 production of baby Swiss cheese, and to evaluate the impact of diet on cow lactation
32 performance. Lactating Holstein dairy cows were fed both dietary treatments in a 2 × 2 crossover
33 design. Cows were housed in a 48-cow free stall pen equipped with Calan gates to record
34 individual feed intake. Diet one (**control**) was a corn/corn silage/hay diet supplemented with
35 SoyPlus®. Diet two was the same base ration, but 20% (DM-basis) reduced-fat dried distillers
36 grains with solubles (**RF-DDGS**) were included in place of SoyPlus®. The RF-DDGS diet was

37 additionally supplemented with rumen protected lysine; diets were formulated to be isoenergetic
38 and isonitrogenous. Cows were allowed *ad libitum* access to feed and water, fed twice daily, and
39 milked three times daily. For cheese production, milk was collected and pooled six times for
40 each dietary treatment. There was no treatment effect on milk yield (35.66 and 35.39 kg/day,
41 control and RF-DDGS, respectively), milk fat production (1.27 and 1.25 kg/day), milk fat
42 percentage (3.65 and 3.61%), milk protein production (1.05 and 1.08 kg/day), lactose percentage
43 (4.62 and 4.64%), milk total solids (12.19 and 12.28%), and somatic cell count (232.57 and
44 287.22×10^3 cells/mL). Milk protein percentage, however, was increased (3.01 and 3.11%) and
45 milk urea nitrogen decreased (14.18 and 12.99 mg/dL), indicating that protein utilization may be
46 more efficient when cows are fed RF-DDGS supplemented with lysine. Additionally, DMI and
47 milk yield were unaffected by treatment. Consequently, there was no treatment effect on any
48 measures of feed efficiency. No differences in cheese were found by a trained panel, except
49 cheese appearance; control cheese eyes were significantly (but not practically) larger than the
50 RF-DDGS cheese. These results indicate that RF-DDGS with supplemental lysine can be
51 effectively utilized in the rations of lactating Holstein dairy cows without negatively influencing
52 production parameters or the physiology of the cow. Finally, feeding RF-DDGS did not
53 negatively influence quality and suitability of milk for production of baby Swiss cheese.

54

55 **Keywords:** efficiency, DDGS, milk fat, protein utilization, sensory

INTRODUCTION

56

57 Feeding of traditional (full-fat) corn dried and wet distillers grains with solubles (**DGs**) to
58 ruminant animals has been studied and reviewed extensively (Klopfenstein et al., 2008;
59 Schingoethe et al., 2009). However, results are mixed, with some studies reporting no changes
60 in production parameters (Anderson et al., 2006; Sasikala-Appukuttan et al., 2008) and others
61 reporting altered milk composition or yield, either positively or negatively, when DG are fed
62 (Kleinschmit et al., 2006; Abdelqader et al., 2009). One problem with feeding traditional DGs is
63 that they contain approximately 13% fat, which is composed of mainly unsaturated fatty acids.
64 This problem is multi-faceted; inclusion of DGs can result in diets that contain concentrations of
65 fat that exceed five percent, which can inhibit fiber digestion (Zinn, 1989). Secondly, problems
66 can arise relating to the great concentration of unsaturated fatty acids in DGs that remain after
67 the fermentation process. Unsaturated fatty acids are toxic to rumen microbes (Maia et al., 2007)
68 and undergo biohydrogenation in the rumen because the rumen has a great capacity for reducing
69 unsaturated hydrocarbons. Incomplete biohydrogenation of these hydrocarbons, however, can
70 result in the production of bioactive forms of conjugated linoleic acid (*trans*-10 *cis*-12 CLA),
71 which inhibit *de novo* lipogenesis in the mammary gland (Baumgard et al., 2001). The
72 aforementioned problem can result in milk fat depression, that is characterized by decreased milk
73 fat without concomitant alteration in concentrations of other milk components (Bauman and
74 Griinari, 2001). With the relatively recent improvements in oil extraction from DGs (Majoni et
75 al., 2011) because of the economic value of the corn oil extracted, traditional high fat DGs
76 probably will be less available in the future. Indeed, personal conversations with regional
77 farmers have indicated that reduced-fat dried distillers grains with solubles (**RF-DDGS**; ~6% fat)
78 are typically the only form of DGs available. While the lower energy content of RF-DDGS is a

79 concern for producers of monogastric animals, it could be an advantage for dairy producers by
80 allowing them to include greater concentrations of this typically economical protein source.
81 Research has shown positive results of lactation performance when RF-DDGS are included in
82 the rations of lactating dairy cows (Mjoun et al., 2010; Castillo-Lopez et. al., 2014; Ramirez-
83 Ramirez et al., 2016).

84
85 Additionally, there are mixed reports about the impact of feeding DGs on cheese quality (Houck
86 et al., 2007; Manimanna Sankarlal et al., 2015). The inclusion of DDGS in the rations of
87 lactating dairy cows has been implicated as a cause of late blowing defects both by the scientific
88 community and the dairy community, including dairy producers (Personal conversation; Houck
89 et al., 2007). Our research group investigated the effects on the quality of baby Swiss cheese of
90 feeding full-fat DDGS to dairy cows and found no differences in the quality of baby Swiss
91 cheese when cows were fed a conventional total mixed ration (TMR) or a TMR containing full-
92 fat DDGS (~ 13% fat; Manimanna Sankarlal et al., 2015).

93
94 To extend our DDGS research, our objectives were to investigate the effects of feeding RF-
95 DDGS (~ 6% fat) to lactating Holstein dairy cows on feed efficiency, the composition of milk
96 produced, and the quality of baby Swiss cheese produced from that milk compared with a control
97 TMR that does not include RF-DDGS. On the basis of our previous research, we hypothesize
98 that feeding RF-DDGS to lactating Holstein dairy cows will not adversely affect feed efficiency,
99 milk composition, or quality of baby Swiss cheese made from the milk of those cows.

100

101
102
103
104
105
106
107
108
109
110
111
112
113
114
115
116
117
118
119
120
121
122
123

MATERIALS AND METHODS

Animals and Diets

All procedures were approved by the Iowa State University Animal Care and Use Committee (IACUC). Thirty-six multiparous mid-lactation Holstein dairy cows (body weight 680 ± 11 kg, 52.25 ± 27.34 DIM) were assigned to one of two dietary treatment groups ($n = 18$ per group) in a 2×2 crossover design. Each experimental period lasted 35 days, with the first 14-d used as an acclimation period. Rations were formulated to meet NRC requirements, to be isonitrogenous and isoenergetic (Tables 1 and 2), and to contain similar intestinally available amino acid concentrations. Ration one (control) was a corn/corn silage/hay based TMR supplemented with SoyPlus® (Dairy Nutrition Plus, Ralston, IA) as a protein source. Ration two was formulated by using the same base ration as the control but with 20% of the dry matter being RF-DDGS (Poet Biorefining, Jewell, IA) containing approximately 6.0% fat in place of SoyPlus® (Table 1). The RF-DDGS ration was supplemented with rumen-protected lysine (Kemin, Des Moines, IA) to make diets similar in available lysine (Table 1). The cows were fed individually by using Calan gates (American Calan, Northwood, NH), allowing for measurement of individual feed intake. Cows were housed together at the Iowa State University Dairy Farm (Ames, IA) in a 48-cow, free-stall pen and individually fed twice daily (0800 h and 1600 h) to allow for approximately 10% refusal. Refusals were weighed daily and recorded. Feed ingredients in the TMR were mixed by using a Patz V615 mixer (Patz Corporation, Pound, WI). Cows were allowed *ad libitum* access to food and water, except during their three daily milkings (8 h apart). Initially, cows were allowed to adapt to using the Calan gates before start of the acclimation period. Additionally, individual milk production was recorded daily by using a Boumatic milking system (Boumatic LLC, Madison, WI).

124

125 *Sample Collection and Analyses*

126 Because the first 14-d of each experimental period were an acclimation period, those data were
127 excluded from analyses. Feed samples were collected three times per experimental period after
128 the acclimation period, and proximate analyses were done by wet chemistry (Dairylands Lab,
129 Arcadia, WI). Fiber (acid detergent) was quantified by AOAC Official Method 973.18 (1996)
130 and lignin by AOAC Official Method 973.18; ether extract was determined by using AOAC
131 Official Method 945.16 and AOAC Official Method 920.39; feed fatty acids were quantified by
132 using the method described by Sukhija and Palmquist (1988); nitrogen was quantified by using
133 AOAC Official Method 990.0; minerals were determined by ICP-MS by using AOAC Official
134 Method 985.0 and AOAC Official Method 2011.14; NDF was determined as described by
135 Mertens (2002); AD-ICP was determined by using AOAC Official Method 973.18 and AOAC
136 Official Method 990.03; ash was determined by AOAC Official Method 942.05; dry matter was
137 determined by using NFTA Method 2.1.4. After the 14-d acclimation period, individual milk
138 samples were collected weekly from all three milkings to represent a 24-h lactation period.
139 Individual milk samples (three from each test day) were sent for proximate analyses and assay of
140 milk urea nitrogen and somatic cell counts (Dairy Lab Services, Dubuque, IA).

141

142 Blood samples were collected during the final week of each period by jugular venipuncture and
143 placed into lithium heparinized vacutainers for NEFA quantification and into fluoridated
144 vacutainers for blood glucose analysis (Becton, Dickson, and Company, Franklin Lakes, NJ).
145 Blood glucose was assayed by following the manufacturer's protocols (Wako Autokit Glucose,
146 Wako Diagnostics, Richmond, VA). Blood NEFA were quantified by following manufacturer's

147 protocol (Wako HR Series NEFA-HR, Wako Diagnostics, Richmond, VA). Rumen fluid was
148 collected via esophageal tube approximately four h post-feeding coinciding with maximal
149 volatile fatty acid (VFA) production. The first few hundred milliliters of rumen fluid were
150 discarded to limit salivary contamination. The next volume of rumen fluid was strained through
151 cheese cloth and frozen at -20°C. Rumen pH was measured immediately on-farm after
152 collection.

153 *Cheese Making*

154 During each of the three 35-d periods, milk from one complete milking (collected at
155 approximately 0630 h) of each treatment group (control or RF-DDGS) was collected for cheese
156 making two to three times during weeks three and four (n = 14 cheeses). The milk cans and
157 dump buckets were washed with automatically diluted Ecolab® Oasis Enforce (St. Paul, MN)
158 and sanitized with automatically diluted Ecolab® Mikroklene® (St. Paul, MN) iodine-based
159 sanitizer. Teats were sanitized with 1000 ppm chlorine predip (ECAcept technology, Zurex
160 PharmAgra LLC, Middleton, WI) and wiped dry with individual towels before collecting milk
161 from each cow by the Boumatic milking system (Boumatic, Madison, WI). Milk from two
162 groups of six cows was collected by re-routing the Boumatic line into a clean dump bucket.
163 After the milk of two cows fed the same diet filled a dump bucket, it was dumped through
164 cheesecloth into a labeled milk can. Milk was transported at ambient temperature to the ISU
165 Center for Crops Utilization Research (CCUR) pilot plant in the Food Sciences Building at Iowa
166 State University (Ames, IA) within 20 min of collection of milk from the last cow. The milk cans
167 were immediately weighed and tested for fat, protein, and lactose prior to further processing
168 (within 60 min of collection) by using a LactiCheck Milk Mini Analyzer (Page and Pederson Inc,
169 Hopkinton, MA). Those who collected milk at the dairy farm showered and changed into clean

170 clothes before participation in cheese making to minimize additional external contamination of
171 milk to be used for cheese production.

172

173 If the fat:protein ratio was not 0.88 ± 0.05 , the milk was separated and standardized; cream or
174 skim from the milk collected from the same experimental cows was added to raise or lower the
175 ratio to 0.88 ± 0.05 , respectively. Milk was separated by using a Type LWA 205 Westfalia
176 Separator (219 rpm in 2.5 dial setting, Dusseldorf, Germany). Approximately 68 kg of
177 standardized milk from each dietary treatment was poured into a labeled cheese vat and heat-
178 treated (63°C for 2 min, to decrease the initial load of bacteria) by delivering steam-heated water
179 to the jacketed vat while the milk was gently agitated. After heat-treatment, the milk was
180 gradually cooled to 33°C by running cold water in the jacketed vat while the milk was gently
181 agitated.

182

183 Baby Swiss-style cheese was made by adding 0.32g (± 0.03) CHOOZIT TA 60 (*Streptococcus*
184 *salivarius* ssp. *thermophilus*, DuPont™ Danisco®, New Century, KS) and 0.12 g (± 0.02)
185 CHOOZIT eyes (*Propionibacterium freudenfeichii* ssp. *shermanii*, DuPont™ Danisco®) per 100
186 kg of 33°C milk. Microbial coagulant (13 mL/100 kg of milk, DCI Supreme, Dairy Connection
187 Inc., Madison, WI) was diluted with cold water to a ratio of 1:40 and added with slow agitation
188 for 1 min after 30 min of ripening. The cheese curd was allowed to set for approximately 30-
189 min, tested for firmness manually and visually, and manually cut with 12-cm wire curd knives.
190 After a 5-min heal, the 35-min forework period of intermittent gentle stirring occurred. About
191 25% of the vat volume of whey was removed (pre-draw), followed by constant stirring and
192 addition of water (3 to 5% of the vat volume) at 33°C . Gradually, the curds were cooked by

193 increasing the temperature to 40°C over a 15-min period, and then to 46°C over a 10-min period
194 by adding steam to the jacket of the vat, with continuous curd stirring. Warm water (~10 % of
195 the vat volume) was added at 44°C to facilitate the rise in temperature of the cheese to
196 46°C(±1°C), at which point the curds were held for 42 min (postwork). Whey was removed
197 after postwork at a target pH of 6.4.

198

199 Approximately 4 h from the time culture was added to the milk, pressing began. Cheese curds
200 were collected into perforated stainless steel Longhorn hoops and pressed, in the whey, by using
201 a 7 kg weight (0.02 kg/cm²), on each hoop follower for 15 min. The whey was drained
202 completely and the cheese block was pressed for 1 hr with 11 kg (0.04 kg/cm²), 1 hr with 23 kg
203 (0.07 kg/cm², and an additional 3 h with 35 kg of weights (0.11 kg/cm²). The weights were
204 removed, and cheese was ripened in an empty basin for an additional 5 to 8 h at 22°C ± 3°C.
205 The ripening time was based on the time required for the pH of the cheese to drop to 5.25
206 (±0.05) (Accumet® Basic AB15, Fisher Scientific Inc, Pittsburgh, PA). According to
207 Kosikowski and Mistry (1997), the optimal pH at the time of salting for Swiss-style cheeses is
208 between 5.2 and 5.4. Cheese rounds were removed from the hoops, cut in half, and placed in
209 saturated brine containing 23% NaCl and 0.38% CaCl₂, for up to 7 to 9 hr (depending on block
210 weight, which averaged 5.3 kg; approximately 30 min/kg cheese).

211

212 Cheese surfaces were allowed to dry for 1 h before rounds were vacuum-packed in clear vacuum
213 seal bags (12 X 16 3-mil; UltraSource® LLC, Kansas City, MO) with a Koch vacuum packing
214 machine (Koch Equipment LLC©, Kansas City, MO). Cheeses were stored at 10 ± 1°C for 7 d

215 (pre-cool), $22 \pm 3^{\circ}\text{C}$ for 21 d (warm room), and $4 \pm 1^{\circ}\text{C}$ (cold room) for 60 d (Kosikowski and
216 Mistry, 1997). Cheeses were flipped weekly.

217

218 *Cheese Analyses*

219 Proximate analysis was conducted by using standard methodology, at South Dakota State
220 University. Fat was analyzed by Mojonnier method (Atherton and Newlander, 1977), moisture
221 content by using a forced-draft oven (model OV-490A-2; Blue M, Blue Island, IL), chloride by
222 using a Corning Chloride Analyzer (Ciba Corning Diagnostics, Medfield, MA), based on the
223 Volhard test (Marshall, 1992 added), and total protein by measuring total nitrogen in the cheeses
224 using the by Kjeldahl method (Kjeltec™ 2200 Auto Distillation Unit, FOSS, Eden Prairie, MN).

225

226 *Sensory Analyses*

227 Sensory quality evaluation began after 62 ± 6 d aging. A descriptive sensory analysis panel,
228 composed of six trained panelists, evaluated the quality of the cheeses. Procedures for use of
229 human subjects in research were approved by the Iowa State University Institutional Review
230 Board (Ames, IA). Training consisted of at least 5 h of initial training, followed by an additional
231 hour of refresher training between the first and second official tasting periods (separated by a
232 month). The expectations for “ideal” characteristics of Swiss and baby Swiss cheese were
233 explained to panelists (Cakir and Clark, 2009). For the appearance attributes, photographs of
234 ideal Swiss and baby Swiss cheeses, as well as those exhibiting varying degrees of specific
235 defects, were used to assist with initial training. To instill specific appearance, aroma, flavor,
236 and body and texture defects in the minds of the panelists, products exhibiting varying levels of
237 defects were utilized, which served as references on a 15-cm unstructured line scale during

238 training sessions (Table 3). The facilitator and panelists created a “Cheat Sheet” to augment
239 training and tasting sessions. The descriptors and references (Table 3) were anchored on the 15-
240 cm unstructured line scale to remind panelists of intensity references. Additionally, to assist with
241 eye size evaluation, panelists were provided a plastic square with a standard hole punched in it to
242 indicate ideal (“small”) eyes and a penny to indicate “large” eyes.

243
244 To representatively sample cheeses for evaluation by panelists, every cheese was systematically
245 cut manually with a sanitized butcher knife on a sanitized cutting board, into at least 20 pieces of
246 approximately 1 cm thickness. The cutter began by making a 1 cm slice at the outside round of
247 the cheese and proceeded to make subsequent slices around the round, gradually forming a
248 smaller and smaller square out of the cheese (Figure 1). Slices were laid out sequentially on
249 sanitized, dry trays for photographing (Figures 2 – 5). When the length, width, and height of the
250 cheese were nearly equal the center-most piece of cheese was flipped vertically and the
251 remaining pieces were cut, resulting in three to four horizontal slices representing the top and
252 bottom “faces” and one to two inner-most slices. Based upon their sequential placement on
253 trays, cheese slices were selected randomly for bagging and presentation to panelists by using a
254 random number generator. A plastic index card-sized template (8.5 cm × 5 cm × 1 cm) was
255 placed on each randomly selected master slice to make a consistent “principal display” for
256 panelist evaluation. Selected slices were placed into individual re-sealable snack bags, labeled
257 with random 3-digit codes corresponding to the original cheese from which they were cut, and
258 stored at 4°C until the sensory panel.

259

260 During actual cheese sensory evaluation sessions, panelists were provided one blank score sheet
261 per sample which was the same as the Cheat Sheet minus the anchored reference words.
262 Panelists were trained to identify appearance, aroma, flavor, and body and texture attributes, but
263 trigger words on the Cheat Sheets served to remind them where on the score sheet to mark the
264 intensity of each cheese attribute on the 15-cm line scale. While in individual booths, panelists
265 were provided individual bagged samples, along with a plastic knife for cutting cheese for body
266 and texture and taste evaluation, as well as water and green grapes for cleansing the palate.
267 Panelists were instructed to first evaluate visual attributes only on the principal display presented
268 to them in the bag. Panelists then evaluated body and texture and finally aroma and flavor.

269

270 *Statistical Analyses*

271 Milk components, yield, and performance metrics were analyzed as a 2×2 crossover design.
272 Data were analyzed by using the MIXED procedure of SAS version 9.4 (Cary, NC). The model
273 included the fixed-effects of treatment, treatment sequence, and period, and the random-effect
274 was cow nested within group. Means with significant treatment effects were separated by using
275 LSMEANS with the PDIFF option. Feed fatty acids and feed proximate analysis results were
276 analyzed by using the MIXED procedure of SAS with the fixed-effect of treatment. Sensory
277 attributes of cheese were analyzed by using the MIXED procedure of SAS with the model
278 including the fixed-effect of treatment. Means were separated by using LSMEANS with the
279 PDIFF option. Differences were considered significant when $P < 0.05$.

280

281

RESULTS AND DISCUSSION

282 As designed, diets were isoenergetic and isonitrogenous (Table 2) as evidenced by the similar
283 NE_L, protein, and total digestible nutrients. Significant differences in phosphorus, sulfur, and
284 calcium resulted in a significantly lower dietary cation anion difference (**DCAD**) in RF-DDGS
285 rations (Table 2). Feed fatty acid compositions are reported in Table 3, and, as expected, the RF-
286 DDGS diet contained slightly greater concentrations of total unsaturated fatty acids because corn
287 oil is 92% unsaturated fatty acids (Ramos et al., 2009).

288

289 Dry matter intake (**DMI**) did not vary significantly between treatment groups, nor did milk yield,
290 fat-corrected milk yield (**FCM**), or energy-corrected milk yield (**ECM**) (Table 4), which is
291 consistent with results of Paz and Kononoff (2014) and Ramirez-Ramirez et al. (2016).

292 Additionally, milk fat percentage (consistent with Ramirez-Ramirez et al., 2016), total milk fat
293 production, total milk protein production, lactose percentage, and total lactose production were
294 unaffected by treatment, which is consistent with results of Paz and Kononoff (2014) and Mjoun
295 et al. (2010). Previous studies have indicated that increased DCAD values can increase milk fat
296 percentage (Harrison et al., 2012; Hu et al., 2007; Wildman et al., 2007); this result, however,
297 was not seen in our study (Table 4), which is consistent with results reported by Erdman et al.
298 (2011). No differences were found in composition of milk from cows fed either diet, with the
299 exception of milk protein percentage, which increased for cows fed RF-DDGS without a
300 concomitant change in total milk protein production. Together, a decrease in milk urea nitrogen
301 (**MUN**) and increased milk protein percentage for cows fed RF-DDGS indicate an increased
302 efficiency of protein extraction from feed (Table 5). These results are consistent with those of

303 Castillo-Lopez et al. (2014) who found that feeding cows RF-DDGS had no effect on milk yield
304 and tended to increase milk protein percentage. Additionally, Ramirez-Ramirez et al. (2016)
305 found that cows fed RF-DDGS produced milk with greater protein percentage and no effect on
306 milk fat percentage, which is consistent with our study. They, however, observed increased milk
307 yield for those cows fed either DDGS or RF-DDGS which was not seen in this study. In this
308 study, milk protein percentage was significantly increased and MUN was decreased when cows
309 were fed RF-DDGS, suggesting that protein utilization may be improved when cows are fed RF-
310 DDGS when supplemented with rumen-protected lysine, which has been reported by Mjoun et
311 al. (2010). Increased milk protein percentage when cows were fed RF-DDGS has been
312 previously reported (Castillo-Lopez et al., 2014; Mjoun et al., 2010; Ramirez-Ramirez et al.,
313 2016). Further research, however, would need to be done to confirm if the improved protein
314 utilization reported in this present study are explained by the observations of Mjoun et al. (2010).
315 Additionally, all metrics of feed efficiency stated in Table 4 were unaffected, which is consistent
316 with prior research that saw no change in feed efficiency when cows were fed RF-DDGS in
317 place of a soy-based protein (Mjoun et al., 2010). Finally, our results mirror those reported by
318 Mjoun et al. (2010), who saw no effect on milk yield, milk fat percentage, and lactose percentage
319 but saw increased protein percentage, and decreased milk urea nitrogen.

320

321 As expected, no difference in body weight change was seen for either treatment group (Table 5)
322 which was reported by Mjoun et al. (2010) as well. In addition, no difference in rumen pH was
323 observed which is consistent with results from trials with similar diets reported by Castillo-
324 Lopez et al. (2014). Furthermore, acetate-to-propionate ratios were decreased and valerate
325 concentration was increased (Table 6), which is consistent with results of Ramirez-Ramirez et al.

326 (2016) who fed similar diets. The increased propionate concentration could help to explain the
327 lack of treatment difference in both lactose concentration, lactose yield, and milk yield. In
328 contrast with the results related to NEFA concentrations of Mjoun et al (2010), we saw no
329 difference in NEFA concentration, which is likely because the cows used in that study were in
330 early lactation, whereas ours were in mid-lactation. Finally, consistent with Mjoun et al. (2010),
331 blood glucose concentrations were not different between treatments. Combined, our results are a
332 confirmation of prior research that reports the effectiveness of including RF-DDGS in the rations
333 of lactating dairy cows (Castilo-Lopez et al., 2014; Mjoun et al., 2010; Ramirez-Ramirez et al.,
334 2016). Furthermore, these results demonstrate that currently commercially available RF-DDGS
335 can be fed at a 20% (DM-basis) inclusion rate without negatively influencing cow performance.
336 Feeding RF-DDGS is especially promising because in this experiment cows fed RF-DDGS
337 performed equally well with those fed the more expensive protein source (SoyPlus®) which
338 offers an economic advantage for the inclusion of RF-DDGS in the rations of lactating dairy
339 cows.

340

341 Swiss cheese and baby Swiss cheese standards for quality evaluation were followed (Cakir and
342 Clark, 2009). Baby Swiss cheese eyes should be glossy, completely round, from 0.3 to 0.8 cm in
343 diameter, and evenly distributed throughout the body of the cheese; the cheese should have a
344 mild nutty (roasted hazelnut) and propionic acid aroma and flavor character with little to no
345 apparent sour/lactic-acid taste. The body of baby Swiss should firm as indicated by being
346 somewhat resistant to initial compression between the thumb, forefinger, and middle finger, but
347 should break apart between fingers without crumbling or seeming too rubbery or dry (corky).
348 Upon mastication, the texture should be smooth rather than grainy or rough. Other than a slight

349 bitter aftertaste, baby Swiss cheese should “clean up”, leaving no fruity, fermented, rancid,
350 yeasty, or other foreign flavors on the palate (Cakir and Clark, 2009).
351
352 Regarding flavor and body and texture attributes of baby Swiss cheese, “ideal” or “typical” mean
353 scores for the attributes of importance would be close to zero (0) for acid, flat, unclean,
354 mealy/grainy, pasty, and weak; the ideal mean score for bitter would be up to 5 and for curdy
355 would be close to 7.5 (Table 6). Mean scores for RF-DDGS and control cheeses were close to
356 the ideal, and not significantly different, for all attributes (Table 7).
357
358 In the case of the present work, ideal mean scores for the attributes of importance would be close
359 to zero (0) for eye distribution, eye shape, and gas formation (defects) and close to the mid-point
360 on the 15-cm line scale (7.5) for eye amount, gloss, and size. The visual appearance of Swiss-
361 type cheeses is very important, and producing cheeses with a typical number, size, and
362 distribution of eyes is difficult (Guggisberg et al. 2015). The baby Swiss cheeses from the
363 control and RF-DDGS treatments in the present study (Table 7 and Figures 2 to 5) had glossy
364 eyes (7.10 and 7.14, respectively), with slightly uneven distribution (3.65 and 3.33, respectively)
365 and a propensity for irregular (somewhat flattened or oval) shape (8.39 and 8.19, respectively).
366 In general, the cheeses looked more like Lorraine® or Lacey Swiss than baby Swiss because the
367 small to very small eyes (5.92 and 5.19, respectively) were overset (7.74 and 7.83, respectively).
368 The RF-DDGS cheeses did not differ from control for any attribute except size of eyes (Table 7).
369 Although mean score for eye size of control cheeses were closer to ideal than that of DDGS
370 cheeses ($P < 0.05$), the difference was not great enough to be practical or obvious to the average
371 consumer. After completion of our research, O’Sullivan et al. (2016) reported on the impact of

372 intentionally adding or not adding *L. helveticus* and *L. casei* to a Swiss-type cheese. In the
373 absence of *L. helveticus*, excess lactose, galactose, and citrate, available during the initial
374 ripening stages, provided substrate to the facultative heterofermentative *L. casei* to produce
375 excessive gas during ripening (overset). Although the presence of individual microorganisms
376 was not enumerated in the present study and sanitation practices were attended to in order to
377 minimize nonstarter bacteria introduction, the absence of *L. helveticus* in the present work may
378 partially explain the overset cheeses, providing support for the work of O'Sullivan et al. (2016).

379
380 Written-in comments by trained panelists and enumeration of appearance defects on 3-digit
381 coded trays of sliced cheeses (representative images included in Figures 2 – 5) support the results
382 compiled in Table 7. When all slices were considered (in addition to the randomly selected
383 samples presented to panelists), uneven eye distribution (84 and 63% of slices from control and
384 RF-DDGS cheeses, respectively), irregular eye shape (76 and 68%, respectively), and very small
385 eyes (59 and 60%, respectively) predominated slices. Blind areas (20 and 17%, respectively),
386 wet/whey (10 and 14%, respectively), and cabbage defects (12 and 8%, respectively) were
387 occasionally seen. Collapsed eyes (20 and 44%, respectively) and slits (18 and 28%,
388 respectively) appeared with higher frequency in RF-DDGS cheeses, but the more serious defect,
389 blown, was more prevalent in control cheeses (33 and 13%, respectively). None of the cheeses
390 were split. Splits and late-blowing defects have a negative impact for both processors
391 (sliceability is affected negatively) and consumers (who expect round glossy eyes); therefore,
392 bad eyes are an economic issue (White et al. 2003; Manimanna Sankarlal et al. 2015). ~~White et~~
393 ~~al. (2003) demonstrated that the propensity for split defects in Swiss cheese was higher in~~
394 ~~summer than winter months and the number of splits in cheese increased with cheese storage~~

395 ~~time. The findings by White et al. (2003) contradicted those of Fröhlich-Wyder et al. (2002),~~
396 ~~who reported that milk produced during the winter hay feeding season was more prone to late~~
397 ~~fermentation defects than summer feeding.~~ To limit late fermentation defects in Swiss-type
398 cheese, Fröhlich-Wyder et al. (2002) recommended propionibacteria strains of low aspartase
399 activity, addition of *L. casei* strains, and omission of *L. helveticus*. To minimize splits White et
400 al. (2003) concluded that careful selection of *L. helveticus* and *P. shermanii* cultures is important
401 particularly if the cheese target moisture content is high (e.g., over 37%). ~~Our work was~~
402 ~~conducted in the winter months,~~ We did not inoculate with *L. helveticus* and cheeses were
403 ripened 78 days (+/- 5) total from brine to knife before evaluation but we did not see any splits.
404 Though we did witness some defects associated with late fermentation, including slits and blow
405 holes, when looked at as a whole, feeding RF-DDGS did not increase the propensity for defects.
406 The findings in the present study are consistent with our previous research (Manimanna
407 Sankarlal et al., 2015) where feeding full-fat DDGS did not affect the quality of baby Swiss
408 cheese produced when compared with feeding a conventional dairy ration without DDGS to
409 lactating Holstein dairy cows.

410

411

CONCLUSION

412 These results indicate that 20% RF-DDGS on a dry-matter basis can be effectively included in
413 the rations of lactating dairy cows without any adverse effects on feed efficiency, milk
414 composition, or measured blood markers of energy balance. These findings indicate promising
415 prospects for the utilization of RF-DDGS, which are often more economical than traditional
416 protein sources. Finally, consistent with previous research (Mjoun et al., 2010), our results
417 indicate that feeding RF-DDGS may lead to improved dietary protein utilization when compared

418 with soybean-based protein. Furthermore, RF-DDGS can be fed without compromising the
419 composition of milk with the exception of slightly increased protein percentage and decreased
420 milk urea nitrogen. Additionally, there were no differences in suitability of milk used for baby
421 Swiss cheese because there was practically no difference in quality of cheese produced from
422 milk from cows conventionally-fed cows and fed RF-DDGS.

423

424

ACKNOWLEDGEMENTS

425 We are grateful for the funding provided by the Minnesota Corn Growers Association and
426 Midwest Dairy Association for supporting this research. In addition, we are thankful to the
427 personnel at the Iowa State University Dairy Farm for their assistance in coordination and
428 implementation of the logistics of performing the work described herein. We are particularly
429 grateful to the many undergraduate assistants who provided labor and care of the research
430 animals, sample collection, and cheese making. We appreciate Dr. Leo Timms for technical
431 assistance on the project and Cristina Duran and Soi Meng Lei for their assistance in the
432 analytical portions of this research. Cultures were generously donated by Dupont™ Danisco®.
433 Proximate analysis of cheese was graciously conducted at South Dakota State University. We
434 also thank Amber Testroet for her assistance in preparation of this manuscript. Finally, we
435 acknowledge the time and expertise provided by our trained panelists.

436
437
438
439
440
441
442
443
444
445
446
447
448
449
450
451
452
453
454
455
456
457
458
459
460
461
462
463
464
465

REFERENCES

Anderson, J. L., D. J. Schingoethe, K. F. Kalscheur, and A. R. Hippen. 2006. Evaluation of dried and wet distillers grains included in two concentrations in the diets of lactating dairy cows. *J. Dairy Sci.* 89:3133-3142.

Atherton, H. V. and J. A. Newlander. 1977. *Chemistry and testing of dairy products*, 4th ed. AVI Publishing Co., Westport, Conn. U.S.A.

AOAC International. 2000. *Official Methods of Analysis*. 17th ed. AOAC Int., Gaithersburg, MD.

Abdelqader, M. M., A. R. Hippen, K. F. Kalscheur, D. J. Schingoethe, and A. D. Garcia. 2009. Isolipidic additions of fat from corn germ, corn distillers grains, or corn oil in dairy cow diets. *J. Dairy Sci.* 92:5523-5533.

Bauman, D. E., and J. M. Griinari. 2001. Regulation and nutritional manipulation of milk fat: Low-fat milk syndrome. *Livest. Prod. Sci.* 70:15-29.

Baumgard, L. H., J. K. Sangster, and D. E. Bauman. 2001. Milk fat synthesis in dairy cows is progressively reduced by increasing supplemental amounts of trans-10 cis-12 conjugated linoleic acid (CLA). *J. Nutr.* 131:1764-1769.

Cakir, E., and S. Clark. 2009. *Swiss Cheese and Related Products*. Chapter 14 in: *The Sensory Evaluation of Dairy Products*. S. Clark, M. Costello, M. Drake and F. Bodyfelt, ed. Springer, New York, NY.

Castillo-Lopez, E., H. A. Ramirez-Ramirez, T. J. Klopfenstein, D. Hostetler, K. Karges, S. C. Fernando, and P. J. Kononoff. 2014. Ration formulations containing reduced-fat dried distillers grains with solubles and their effect on lactation performance, rumen fermentation, and intestinal flow of microbial nitrogen in Holstein cows. *J. Dairy Sci.* 97:1578-1593.

Erdman, R. A., S. Piperova, and R. A. Kohn. 2011. Corn silage versus corn silage:alfalfa hay mixtures for dairy cows: Effects of dietary potassium, calcium, and cation-anion difference. *J. Dairy Sci.* 94:5105-5110.

Fröhlich-Wyder, M, H. P. Bachmann, and M. G. Casey. 2002. Interaction between propionibacteria and starter / non-starter lactic acid bacteria in Swiss-type cheeses. *Lait* 82:1-15.

- 466 Guggisberg, D., P. Schuetz, H. Winkler, R. Amrein, E. Jakob, M.-T. Fröhlich-Wyder, S. Irmeler,
467 W. Bisig, I. Jerjen, M. Plamondon, J. Hofmann, A. Flisch, and D. Wechsler. 2015.
468 Mechanism and control of the eye formation in cheese. *Int. Dairy J.* 47:118-127.
- 469 Harrison, J., R. White, R. Kincaid, E. Block, T. Jenkins, and N. St. Pierre. 2012. Effectiveness of
470 potassium carbonate sesquihydrate to increase dietary cation-anion differences in early
471 lactation cows. *J. Dairy Sci.* 95:3919-3925.
- 472 Houck, K. B., W. L. Wendorff, and R. M. Kaiser. 2007. Distillers grains can add nutrition – and
473 spores. Wisconsin Center for Dairy Research. *Dairy Pipeline.* 19:4-6.
- 474 Hu, W., M. R. Murphy, P. D. Constable, and E. Block. 2007. Dietary cation-anion difference and
475 dietary protein effects on performance and acid-base status of dairy cows in early
476 lactation. *J. Dairy Sci.* 90:3355-3366.
- 477 Kleinschmit, D. H., D. J. Schingoethe, K. F. Kalscheur, and A. R. Hippen. 2006. Evaluation of
478 various sources of corn dried distillers grains plus solubles for lactating dairy cattle. *J.*
479 *Dairy Sci.* 89:4784-4794.
- 480 Klopfenstein, T. J., G. E. Erickson, and V. R. Bremer. 2008. Board invited review: Use of
481 distillers by-products in the beef cattle feeding industry. *J. Anim. Sci.* 86:1223-1231.
- 482 Kosikowski, F. V. and V. V. Mistry. 1997. *Cheese and Fermented Milk Foods: Volume II*
483 *Procedures and Analysis.* F. V. Kosikowski, LLC: Westport, CT.
- 484 Maia, M. R. G., L. C. Chaudhary, L R. Figueres, J. R. Wallace. 2006. Metabolism of
485 polyunsaturated fatty acids and their toxicity to the microflora of the rumen. *J. Microbiol.*
486 91:303-314.
- 487 Marshall, R. T. 1992. *Standard Methods for the Examination of Dairy Products*, 16th Edition.
488 American Public Health Association: Washington, DC.
- 489 Majoni, S., T. Wang, and L. A. Johnson. 2011. Physical and chemical processes to enhance oil
490 recovery from condensed corn distillers solubles. *J. Am. Oil Chem. Soc.* 88:425-434.
- 491 Manimanna Sankarlal, V., E. D. Testroet, D. C. Beitz, and S. Clark. 2015. Dried distillers grains
492 with solubles do not always cause late blowing in baby Swiss cheese. *J. Dairy Sci.*
493 98:8545-8553.

494 Mertens, D.R. 2002. Gravimetric determination of amylase treated neutral detergent fiber in
 495 feeds with refluxing in beakers or crucibles: Collaborative Study. J. AOAC Int. 85:1217-
 496 1240.

497 Mjoun, K., K. F. Kalscheur, A. R. Hippen, D. J. Schingoethe. 2010. Performance and amino acid
 498 utilization of early lactation dairy cows fed regular or reduced-fat dried distillers grains
 499 with solubles. J. Dairy Sci. 93:3176-3191.

500 O’Sullivan, D. J., P. L. H. McSweeney, P. D. Cotter, L. Giblin, and J. J. Sheehan. 2016.
 501 Compromised *Lactobacillus helveticus* starter activity in the presence of facultative
 502 heterofermentative *Lactobacillus casei* DPC6987 results in atypical eye formation in
 503 Swiss-type cheese. J. Dairy Sci. 99: 2625-2640.

504 Paz, H. A. and P. J. Kononoff. 2014. Lactation responses and amino acid utilization of dairy
 505 cows fed low-fat distillers dried grains with solubles with or without rumen-protected
 506 lysine supplementation. J. Dairy Sci. 97:6519-6530.

507 Ramos, M. J., C. M. Fernandez, A. Casas, L. Rodriguez, and A. Perez. 2009. Influence of fatty
 508 acid composition of raw materials on biodiesel properties. Bioresour. Tech. 100:261-268.

509 Ramirez-Ramirez, H. A., E. Castillo-Lopez, C. J. R. Jenkins, N. D. Aluthge, C. Anderson, S. C.
 510 Fernando, K. J. Harvatine. P. J. Kononoff. 2016. Reduced-fat dried distillers grains with
 511 solubles reduces the risk for milk fat depression and supports milk production and
 512 ruminal fermentation in dairy cows. J. Dairy Sci. 99:1912-1928

513 Sasikala-Appukuttan, A. K., D. J. Schingoethe, A. R. Hippen, K. F. Kalscheur, K. Karges, and
 514 M. L. Gibson. 2008. The feeding value of corn distillers solubles for lactating dairy cows.
 515 J. Dairy Sci. 91:279-287.

516 Schingoethe, D. J., K. F. Kalscheur, A. R. Hippen, and A. D. Garcia. 2009. Invited review: The
 517 use of distillers products in dairy cattle diets. J. Dairy Sci. 92:5802-5813.

518 Sukhija.P. S.and Palmquist. D. L. 1988. Rapid method of determination of total fatty acid
 519 content in composition of feedstuffs and feces. J. Agric. Food Chem. 36:1202-1206

520 White, S. R., J. R. Broadbent, C. J. Oberg, and D. J. McMahon. 2003. Effect of *Lactobacillus*
 521 *helveticus* and *Propionibacterium fredenrichii* ssp. *shermanii* combinations on propensity
 522 for split defect in Swiss cheese. J. Dairy Sci. 86:719-727.

- 523 Wildman, C. D., J. W. West, and J. K. Bernard. 2007. Effects of dietary cation-anion difference
524 and potassium to sodium ratio on lactation dairy cows in hot weather. J. Dairy Sci.
525 90:970-977.
- 526 Zinn, R. A. 1989. Influence of level and source of dietary fat on its comparative feeding value in
527 finishing diets for feedlot steers. J. Anim. Sci. 67:1038-1049.
- 528 .

529
530
531
532
533
534
535
536
537
538
539
540
541
542
543
544
545

TABLES AND FIGURES

List of figures

Figure 1. Visual representation of the process used to systematically slice cheeses for photographs and evaluation by trained panelists

Figure 2. Representative photograph of a tray of sliced baby Swiss cheese from milk of cows fed control diet in period 1

Figure 3. Representative photograph of a tray of sliced baby Swiss cheese from milk of cows fed RF-DDGS diet in period 1

Figure 4. Representative photograph of a tray of sliced baby Swiss cheese from milk of cows fed control diet in period 2

Figure 5. Representative photograph of a tray of sliced baby Swiss cheese from milk of cows fed RF-DDGS diet in period 2

Table 1. Formulations of control and reduced-fat dried distillers grains with solubles (RF-DDGS) rations

Ingredient, % dry matter	Treatment	
	Control	20% RF-DDGS
Corn silage	35.13	31.57
Alfalfa hay	23.09	20.74
Whole cotton seed	8.03	7.21
Ground corn	14.41	15.13
RF-DDGS	0.00	19.45
SoyPlus® ¹	13.51	0.54
Quality Liquid Feeds ²	3.81	3.42
USA Lysine ³	0.00	0.11
Vitamin and mineral premix	3.81	3.42

547 ¹ Dairy Nutrition Plus, Des Moines, IA.

548 ² Quality Liquid Feeds, Dunlap, IA. Custom vitamin and mineral supplement.

549 ³ Kemin Industries, Des Moines, IA

550

Table 2. Analysis of control and reduced-fat dried distillers grains with solubles (RF-DDGS) rations

Item	Treatment		SEM	P-Values
	Control	20% RF-DDGS		
Moisture, %	43.10	40.83	0.956	0.125
Dry matter (DM), %	56.90	59.17	0.956	0.125
Crude protein, % DM	18.09	17.86	0.213	0.446
ADF ¹ , % DM	21.37	20.52	0.678	0.397
aNDF ² , % DM	28.78	29.35	0.757	0.602
aNDF, % OM ³	27.93	28.20	0.776	0.807
Lignin, % DM	4.01	4.01	0.181	0.985
Lignin, % NDF	14.34	14.22	0.387	0.836
AD-ICP ⁴ , % DM	0.93	1.02	0.070	0.228
ND-ICP ⁵ , % DM	2.90	2.90	0.033	0.434
Fat, % DM	4.80	5.21	0.130	0.052
Ash, % DM	8.60	8.20	0.170	0.117
Calcium, % DM	1.16	0.94	0.052	0.014
Phosphorus, % DM	0.34	0.44	0.013	0.0003
Magnesium, % DM	0.26	0.24	0.016	0.567
Potassium, % DM	1.96	1.81	0.051	0.069
Sulfur, % DM	0.25	0.36	0.010	<0.0001
Sodium, % DM	0.56	0.51	0.016	0.070
Chloride, % DM	0.63	0.63	0.022	0.873
DCAD, mEq/100g	40.81	27.89	1.766	0.0004
TDN ⁶ , 1 × %DM	71.56	72.09	0.505	0.481
NE _L ⁷ , Mcal/kg	1.64	1.65	0.012	0.479

551 ¹Acid-detergent fiber552 ²Neutral-detergent fiber553 ³Organic matter554 ⁴Acid-detergent insoluble crude protein555 ⁵Neutral-detergent insoluble crude protein556 ⁶Total digestible nutrients557 ⁷Net energy for lactation

Table 3. Feed fatty acid composition

Fatty acid, wt%	Treatment			<i>P</i> -Value
	Control	RF-DDGS ¹	SEM	
C12:0, C14:0	0.30	0.23	0.085	0.611
C14:1	0.00	0.00	NA	NA
C16:0	20.27	19.20	0.299	0.029
C16:1	0.37	0.28	0.017	0.006
C17:0	0.00	0.00	NA	NA
C18:0	3.22	2.68	0.080	0.0005
C18:1	18.68	20.55	0.209	<0.0001
C18:2	50.02	51.43	0.265	0.004
C18:3	5.71	4.21	0.147	<0.0001
C19:0, C20:0	0.58	0.60	0.012	0.222
C20:1	0.27	0.30	0.008	0.007
C20:2, C20:3	0.00	0.00	NA	NA
C20:4	0.00	0.00	NA	NA
C22:1	0.00	0.00	NA	NA
C22:6	0.00	0.00	NA	NA
C24:0	0.00	0.00	NA	NA
C24:1	0.00	0.00	NA	NA
tUFA ²	75.04	76.77	0.320	0.003
MUFA	19.31	21.14	0.200	<0.0001
PUFA	55.73	55.63	0.252	0.799

¹Reduced-fat dried distillers grains with solubles

²Total unsaturated fatty acids

559

560

561

Table 4. Effects of feeding RF-DDGS to lactating Holstein dairy cows on milk components and yield

Item	Treatment			
	Control	RF-DDGS	SEM	<i>P</i> -Value
Dry matter intake (DMI), kg/day	20.69	20.89	0.538	0.200
Milk yield, kg/day	35.66	35.39	0.978	0.329
FCM ¹	36.27	35.78	0.889	0.105
ECM ²	36.43	36.30	0.887	0.663
Milk fat, kg/day	1.27	1.25	0.043	0.416
Milk fat, %	3.65	3.61	0.096	0.517
Milk protein, kg/day	1.05	1.08	0.032	0.204
Milk protein, %	3.01	3.11	0.051	0.002
Lactose, kg/day	1.63	1.62	0.057	0.884
Lactose, %	4.62	4.64	0.050	0.819
Milk total solids, %	12.19	12.28	0.167	0.478
Milk urea nitrogen, mg/dL	14.18	12.99	0.285	<0.0001
Somatic cell count/mL × 1000	232.57	287.22	168.84	0.718
Feed efficiency, kg milk/kg DMI	1.78	1.75	0.062	0.168
FCM efficiency, kg FCM/kg DMI	1.81	1.77	0.056	0.110
ECM efficiency, kg ECM/kg DMI	1.82	1.80	0.056	0.351

¹Fat-corrected milk = (0.432 × milk)+(16.23 × milk fat)

²Energy-corrected milk = (0.327 × milk)+(12.95 × milk fat)+(7.65 × milk protein)

562

563

564

Table 5. Effects of feeding RF-DDGS to dairy cows on body weight, blood components, rumen fluid pH and ruminal short-chain fatty acid concentrations

Item	Treatment			
	Control	RF-DDGS	SEM	<i>P</i> -Value
Body weight change, kg	+12.19	+17.33	2.43	0.139
Rumen fluid pH	6.55	6.50	0.057	0.365
Blood NEFA, μ eq/L	164.54	159.53	6.10	0.508
Blood glucose, mg/dL	53.08	54.85	1.24	0.319
Short-chain fatty acids, mM				
Acetate	40.40	41.68	2.40	0.707
Butyrate	7.03	7.80	0.44	0.227
Isobutyrate	0.39	0.42	0.03	0.484
Isovalerate	1.70	1.57	0.11	0.415
Propionate	13.54	16.02	1.11	0.071
Valerate	0.88	1.11	0.06	0.013
Acetate:Propionate	3.16	2.75	0.10	<0.0001

567 **Table 6.** Descriptive analysis lexicon for baby Swiss cheeses (adapted from Cakir and Clark,
568 2009)

Descriptor	Definition and ideal location on 15-cm unstructured line scale	Reference and location on 15-cm scale*
	<i>Aroma, Taste & Flavor</i>	
Acid	A basic taste, characteristic of lactic acid and citric acid Ideal = 0 – 2.5 cm	0 – 2.5 = Skim milk 2.5 – 5 = 1 tsp cultured buttermilk/L milk 5 – 10 = 2 TBS buttermilk/L milk 10 – 15 = cultured buttermilk
Bitter	A basic taste, generally toward the back of the tongue and/or throat, characteristic of quinine or caffeine Ideal = 0 – 5 cm	0 – 2.5 = Skim milk 2.5 – 5 = slight 7.5 – 10 = 1 tsp 0.5% quinine sulfate/L milk 10 – 15 = 1 TBS 0.5% quinine sulfate/L milk
Flat	Practically devoid of characteristic sweet roasted hazelnut flavor of Swiss cheese Ideal = 0 – 2.5 cm	0 – 2.5 = Kraft Big Slice Swiss 2.5 – 5 = slight 5 – 7.5 = definite 10 – 15 = Monterey Jack cheese
Unclean	Persistent, complex and unpleasant flavor; could smell somewhat fecal Ideal = 0 cm	0 – 3.5 = Skim milk 3.5 – 8 = slight 8 – 12 = definite 12 – 15 = Grafton Extra mature Cheddar
	<i>Body & Texture</i>	
Curdy	Resistant to compression between thumb and fore-fingers, slightly rubbery Ideal = 6 – 8.5 cm	0 – 2.5 = Hy-Vee sharp Cheddar 2.5 – 5 = slight 5 – 7.5 = Shullsburg Creamery baby Swiss 7.5 – 10 = definite 10 – 15 = Hy-Vee mild Cheddar
Mealy/grainy	Coarse, crumbly mass between fingers; small sandy to rough granular particulates in mouth with mastication Ideal = 0 – 5 cm	0 – 2.5 = Cream cheese 2.5 – 7.5 = slight 7.5 – 10 = Cacique Cotija cheese 10 – 15 = pronounced
Pasty	Sticky between fingers and in mouth Ideal = 0 – 2.5 cm	0 – 2.5 = not 2.5 – 7.5 = slight 7.5 – 10 = definite 10 – 15 = cream cheese
Weak	Immediately yields between fingers when worked between thumb and fore-fingers Ideal = 0 – 2.5 cm	0 – 2.5 = Shullsburg Creamery baby Swiss 2.5 – 5 = Hy-Vee Monterey Jack 5 – 10 = definite 10 – 15 = Cream cheese
	<i>Appearance</i>	
Eye amount	Blind = no eyes (if seen in one area of the cheese, the cheese is also faulted for uneven distribution)	0 – 2.5 = blind 2.5 – 5 = underset

	Underset = too few eyes (more than 2 cm between eyes) Ideal = 1 to 2 cm distance between eyes Overset = less than 1 cm between eyes, overcrowding Ideal = 5 – 7.5 cm	5 – 7.5 = ideal 7.5 – 10 = slight overset 10 – 12.5 = moderate overset 12.5 – 15 = very overset
Eye distribution	Ideal = even or equal sized spacing between eyes Uneven = tendency for eyes to predominate one area of a cheese, or not be evenly distributed throughout the cheese Ideal = 0 – 2.5 cm	0 – 2.5 = even 2.5 – 6.5 = slight uneven 6.5 – 12 = moderately uneven 12 – 15 = very uneven
Eye gloss	Wet = free whey Ideal = Shiny but not wet Slightly dull = dry, with lack of luster Dull/rough = in extreme cases the inside of an eye may look like a nutshell Ideal = 5 – 7.5 cm	0 – 2.5 = wet 2.5 – 5 = slight wet 5 – 7.5 = Ideal 7.5 – 12 = slight dull 12 – 15 = dull/rough
Eye shape	Ideal = completely round, no defect Irregular = not round (oval, lenticular) Collapsed = eyes appear to have flattened after development Cabbage = multiple adjacent eyes adjoin one-another; a paper-thin layer of cheese may appear between eyes Ideal = 0 – 2.5 cm	0 – 2.5 = no defect 2.5 – 6.5 = few irregular 6.5 – 10 = multiple irregular 10 – 12.5 = collapsed 12.5 – 15 = cabbage
Eye size	Sweet holes = < 0.1 cm diameter Very small = 0.1 to 0.2 cm Small = 0.2 to 0.34 cm Ideal = 0.4 to 0.8 cm diameter; standard punch hole Medium = 0.8 to 1.8 cm diameter Large = >1.8 cm; penny Ideal = 7.5 – 10 cm	0 – 2.5 = sweet holes 2.5 – 5 = very small 5 – 7.5 = small 7.5 – 10 = ideal 10 – 12.5 = medium 12.5 – 15 = large
Gas formation (defect)	Ideal = no defect Checks/picks = tiny (picks, 1-2 mm) to small (checks <5 mm), irregular cracks or ragged openings Slits/splits = slits are cracks (0.5-1.5 cm); splits exceed 1.5 cm and can traverse entire piece of cheese Blow hole = irregular excess gas formation collapses cheese structure, leaving large, irregular gaping hole in cheese Ideal = 0 – 2.5 cm	0 – 2.5 = no defect 2.5 – 5 = picks 5 – 7.5 = checks 7.5 – 10 = slits 10 – 12.5 = splits 12.5 – 15 = blow hole

569 *Photographic images used for appearance references are not included in the present work.

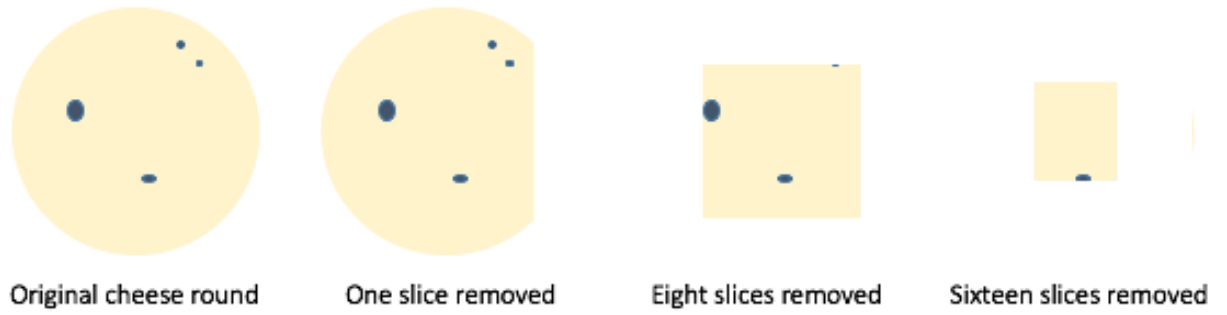
570

Table 7. Mean scores* for cheese flavor, body and texture, and appearance of eyes

Flavor	Treatment		SEM	P - Value
	Control	RF-DDGS		
Acid	0.99	1.07	0.14	0.683
Bitter	3.74	3.48	0.34	0.558
Flat	1.33	1.60	0.23	0.398
Unclean	1.40	1.86	0.29	0.220
Body and Texture				
Curdy	7.88	7.46	0.34	0.361
Mealy/Grainy	5.60	6.44	0.42	0.132
Pasty	0.66	0.91	0.18	0.298
Weak	0.89	1.13	0.16	0.254
Appearance of Eyes				
Amount	7.74	7.83	0.31	0.834
Distribution	3.65	3.33	0.40	0.556
Gloss	7.10	7.14	0.22	0.877
Shape	8.39	8.19	0.28	0.588
Size	5.92	5.19	0.24	0.021
Gas formation (defects)	3.82	4.69	0.59	0.266

571 * Mean values of 14 cheeses, evaluated by a trained descriptive panel of six judges, in duplicate,
572 by using a 15-cm line scale.

573



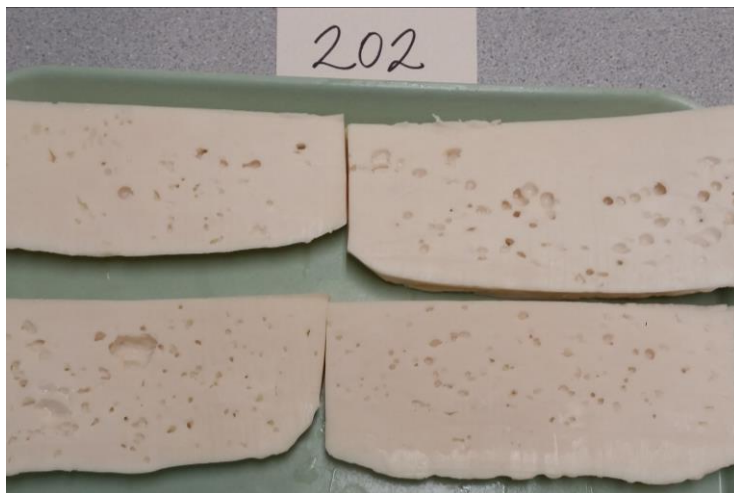
574

575 **Figure 1.**

576

577

578



579

580 **Figure 2.**

581

582



583

584 **Figure 3.**

585

586

587



588

589 **Figure 4.**

590



591

592 **Figure 5.**